

1 **Laboratory Guidance - Drying Field-Fresh Hemp Plant Samples in**

2 **Preparation for Determination of Total THC on a Dry-Weight Basis**

3 AOAC Cannabinoid Analytical Science Program (CASP) Working Group

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7 **1.0 Objective**

8 The objective of this Appendix is to provide general guidance for laboratories to aid in
9 the development of Standard Operating Procedures (SOPs) appropriate for processing hemp
10 plant materials received fresh from the field. Field-fresh materials typically have a higher
11 moisture content of 70-80% (1) requiring a preliminary drying step to reduce moisture content to
12 a value that is sufficiently low (approx. 15%). Dried, milled samples are needed for requisite
13 sample preparation by appropriate methods including AOAC 2018.11 for the determination of
14 total tetrahydrocannabinol (THC) on a dry-weight basis.

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16 **2.0 Purpose and Scope**

17 This document offers guidance to be used by laboratories to aid in the development of
18 procedures to sufficiently dry field-fresh hemp material. The purpose of the procedure is to dry
19 the material enough to facilitate adequate grinding and homogenization prior to analysis. The
20 scope of this guidance is limited to the initial drying step only and is not intended to be used as
21 the primary procedure used for sample preparation for determination and reporting total THC.
22 Complete sample preparation details are described within published methods such as AOAC
23 OMA 2018.11 (2).

24 In this context, the drying procedure needed and discussed herein is a preliminary step
25 that, while representing an important step to ensuring that sample integrity is maintained, is not
26 intended for use to directly generate final, reportable results for total THC (dry-weight basis).

27 It is critical for laboratories to develop a procedure in such a way that it can be applied in
28 a consistent manner to minimize the introduction of measurement error as well as maintain
29 sample integrity. During procedure development and validation, the laboratory should identify,
30 evaluate and document critical steps and parameters of the final procedure, assess overall
31 performance and address details that produce inconsistencies or errors in the final reported
32 results for total THC (dry-wight basis). This document provides guidance with that
33 documentation.

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38 **3.0 Introduction and Background – Drying Hemp Plant Materials**

39 Requirements specified by the United States Department of Agriculture (USDA)
40 Agricultural Marketing Service (AMS) for determination of total THC (dry-weight basis) in
41 hemp explicitly state that hemp plant samples must be delivered to the testing laboratory as
42 harvested from the field, i.e. in field-fresh condition (3). In this state and immediately post-
43 harvest, the material will have a moisture content of between 70 – 80% and, while the USDA
44 AMS rule does not mandate a specific drying process or procedure it does outline that testing
45 laboratories “remove the majority of water” from hemp material prior to milling and
46 homogenizing.

47 This initial drying step is crucial to the removal enough moisture from the plant material
48 received by the laboratory to facilitate effective grinding and homogenization while not changing
49 the material and possibly introducing analytical error. However, this step is not intended to take
50 the material completely to dryness as it will undergo additional sample preparation prior to
51 analysis for the determination of total-THC (dry weight basis). Rather, the target moisture
52 content after this initial drying step is 10-15%. Moisture content significantly higher than this
53 and the material will not mill properly. Moisture content less than 10% can result from overly
54 aggressive drying and lead to change in cannabinoid profile and should also be avoided.

55 Consequently, the sample drying procedure is intended only to be used in preparing field-
56 fresh hemp plant materials for subsequent regulatory analysis that reports total THC through full
57 decarboxylation of tetrahydrocannabinolic acid (THC-A) or using the approved mathematical
58 calculation for total THC shown below in Eq. (1):

59

60

$$\text{Total THC} = 0.877 * \text{THC-A} + \text{delta-9 THC} \quad \text{Eq. (1)}$$

61

62 The drying procedure developed by the laboratory must be carefully crafted in order to
63 minimize compromising the sample and introducing analytical error to the reported total-THC

64 value. There is no singular approach that will be acceptable for use by all laboratories due to a
65 combination of diverse laboratory environmental conditions and available drying equipment.
66 Each laboratory will be required to independently develop, evaluate and verify their own drying
67 procedure.

68 There are three generally recognized mechanisms used for drying plant material: heat, air
69 movement, and dehumidification. Any one or combination of these can and should be considered
70 in the development of the laboratory drying procedure. It should be noted the each of these
71 mechanisms must be well defined and controlled in the final procedure. For example, use of
72 drying temperatures that are too high so as to shorten drying time should be avoided as these
73 may cause decarboxylation of the acid forms of cannabinoids. The combination of drying
74 temperatures that are too high and air movement can volatilize terpenes during the drying
75 process further modifying the cannabinoid profile of the material. Both of these conditions will
76 result in a change to the cannabinoid profile of the material and may introduce unwanted
77 analytical error.

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79 **4.0 Hemp Drying Procedure - Experimental Design Guidance**

80 This section provides guidance on collection of data needed to verify oven temperature
81 and drying times are suitable for the drying of field-fresh hemp to a moisture content of less than
82 15% and appropriate for further laboratory processing prior to analysis. Drying conditions should
83 be verified using procedures that are representative of routine sample processing, with
84 consideration given to oven capacity and include various sample locations within the oven.

85

86 **4.1 Study Protocol Development Considerations**

87 The study Protocol should include all critical procedural details including sample
88 handling, sample weighing, drying container types and handling and specific oven conditions.
89 The Protocol should also include a comprehensive listing of all laboratory equipment needed to
90 support the drying procedure such as types of weighing containers, required scales and balances
91 as well as temperature monitoring equipment and timers. Scales, balances, timers and
92 temperature monitoring equipment should all be calibrated prior to use. Data recording and
93 processing software may also be identified. Customized algorithms developed to support data

94 collection and processing, e.g. Excel, should be evaluated and, if more complex, validated prior
95 to use.

96 To assess the reproducibility of the final drying procedure, various container types (if
97 used in that laboratory) and sample sizes representative of those typically received by the
98 laboratory should be included to evaluate different drying times that may be required.

99 All details included in the study Protocol along with needed calculations should be
100 captured in the final procedure document for laboratory personnel to use. Training, both initial
101 and periodic, is also recommended.

102

103 **4.2 Initial Sample Processing and Sub-sampling**

104 A sample of field-fresh hemp may arrive at the laboratory in varied quantities and in
105 different container types and size. Samples arriving to the laboratory in containers not suitable
106 for drying should be transferred to containers that can withstand drying temperatures and
107 handling conditions. Appropriate containers should be identified during procedure development
108 and can be simple and may include, for example, paper bags. Larger sample quantities that
109 cannot be dried in their original containers may be split and sub-sampled into more manageable
110 sizes. Proper sub-sampling may be required and appropriate procedures must be established to
111 ensure a representative sample is acquired using an established statistical sampling approach
112 (4,5). During procedure development and validation, the impact of any sample handling or sub-
113 sampling on sample integrity, including bias or other analytical error, should be formally
114 assessed and included as a component of expanded measurement uncertainty for reporting total
115 THC.

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117 **4.3 Sample Weighing and Drying Conditions**

118 The recommended drying temperature is $65^{\circ}\text{C} \pm 5^{\circ}\text{C}$ to facilitate reasonable drying times
119 without compromising sample integrity. Using appropriate container type and sample quantity,
120 the weight of the drying container along with the initial weight of the field-fresh sample are
121 measured and recorded. The hemp sample is then placed in/on the weighing container and placed
122 into the drying oven to begin the drying process.

123 During the study, the samples are periodically removed from the oven, weighed with the
 124 weight recorded and returned to the oven for further drying. This process is repeated until a
 125 constant weight is observed. Each weighing during the drying process can be made for the hemp
 126 material alone or a combined weight of the hemp and drying container. The same weighing
 127 scheme selected at the beginning of the drying procedure must be used throughout the drying
 128 process. Drying times should be carefully evaluated. The target moisture content using this
 129 procedure is 10-15% to ensure the material is adequately dry for grinding while still maintaining
 130 sample integrity for determination of Total THC.

131 As samples are withdrawn from the oven and weighed, loss on weighing is calculated
 132 relative to the previous weighing to monitor drying progress. Table 1 provides guidance on how
 133 often to weigh the sample when developing appropriate drying methods.

134

135 *Table 1. Suggested periodicity for collecting weights to determine moisture-loss*
 136 *curve for field-fresh hemp.*

Hemp Sample Weight, g	Weighing Frequency
< 100	6x / Day
100 - 500	3x / Day
> 500	1x / Day

137

138

139 The study should continue with sample weight until at least two successive weight
 140 determinations produce a percent weight change of less than 5%. If samples and containers are
 141 weighed together during the study, percent weight-change, %-Wt Change, is calculated using Eq.
 142 (2) below:

143

$$144 \quad \% \text{ -Wt Change} = (TW_p - TW_c) / (TW_p - W_{\text{cont}}) \times 100 \quad \text{Eq. (2)}$$

145

146 Where:

147 TW_p = Total Weight of hemp sample + container at previous time point, in grams

148 TW_c = Total Weight of hemp sample + container at current time point, in grams

149 W_{cont} = Weight of drying container, in grams

150

151 Note that Eq. (2) requires the weighing of the container used for drying the hemp sample.

152

153 If only sample weights are measured during drying, percent weight-change is calculated using a
154 simplified version of Eq. (2) shown below in Eq. (3):

155

$$156 \quad \% \text{-Wt Change} = (SW_p - SW_c) / (SW_p) \times 100 \quad \text{Eq. (3)}$$

157

158 Where:

159 SW_p = Sample Weight of hemp at previous time point, in grams

160 SW_c = Sample Weight of hemp at current time point, in grams

161

162

163 In addition, manual evaluation of the dried sample should also suggest that the material
164 will readily crumble when handled. If not, the sample is not dry enough for grinding and should
165 be dried for a longer period of time. Larger sample quantities or those with higher moisture
166 content will require more time to dry than smaller, drier sample quantities.

167 Similarly, the total percent moisture removed for a given drying interval relative to the
168 final weighed sample that has reached steady-state can be estimated by using Eq. (4) for sample
169 and drying container weighed together or using Eq. (5) if weighing the sample alone and without
170 including the drying container. As above in Eq (2), Eq (4) requires the weighing of the container
171 used for drying the hemp sample:

172

$$173 \quad \% \text{-Moisture Loss} = (TW_c - TW_f) / (TW_c - W_{\text{cont}}) \times 100 \quad \text{Eq (4)}$$

174

175 Where:

176 TW_c = Total Weight of hemp sample + container at current time point, in grams

177 TW_f = Total Weight of hemp sample + container at final time point, in grams

178 W_{cont} = Weight of drying container, in grams

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$$181 \quad \% \text{-Moisture Loss} = (SW_c - SW_f) / (SW_c) \times 100 \quad \text{Eq (5)}$$

182

183 Where:

184 SW_c = Total Weight of hemp sample at current time point, in grams

185 SW_f = Total Weight of hemp sample at final time point, in grams

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189 **4.4 Example Studies**

190 Data from two example studies are presented in Table 2. The data obtained were
 191 collected for 140 g and 400 g samples of field-fresh hemp oven-dried at 60°C using two different
 192 container types, closed paper bags and a drying pan.
 193

194 *Table 2. Example Data for 140-g and 400-g field-fresh hemp samples oven-dried at*
 195 *60°C in closed paper bags or open pan.*

Time, h	Measured Weight, g	Weight Change ¹ , %	%-Total Moisture Loss ²
----- 140-g Hemp Sample Dried in Paper Bag³ -----			
0	161	-	74
7	117	27	62
18	90	23	46
24	67	26	18
31	61	9	5
42	59	3	0
48	59	0	0
----- 400-g Hemp Sample Dried in Paper Bag³ -----			
0	420	-	76
7	334	22	70
18	254	26	59
24	222	14	52
31	186	18	42
42	158	17	30
48	143	11	21
55	132	9	13
66	121	10	3
72	119	2	1
79	118	1	0
----- 400-g Hemp Sample Dried Using Drying Pan³ -----			
0	932		75
7	820	29	65
18	708	40	41
24	643	39	4
31	640	3	1
42	639	1	0
¹ Percent weight change per Eq. (2) or Eq. (3). ² Percent moisture per Eq. (4) or Eq. (5). ³ Container weights – Paper Bag: 23 g; Drying Pan: 540 g.			

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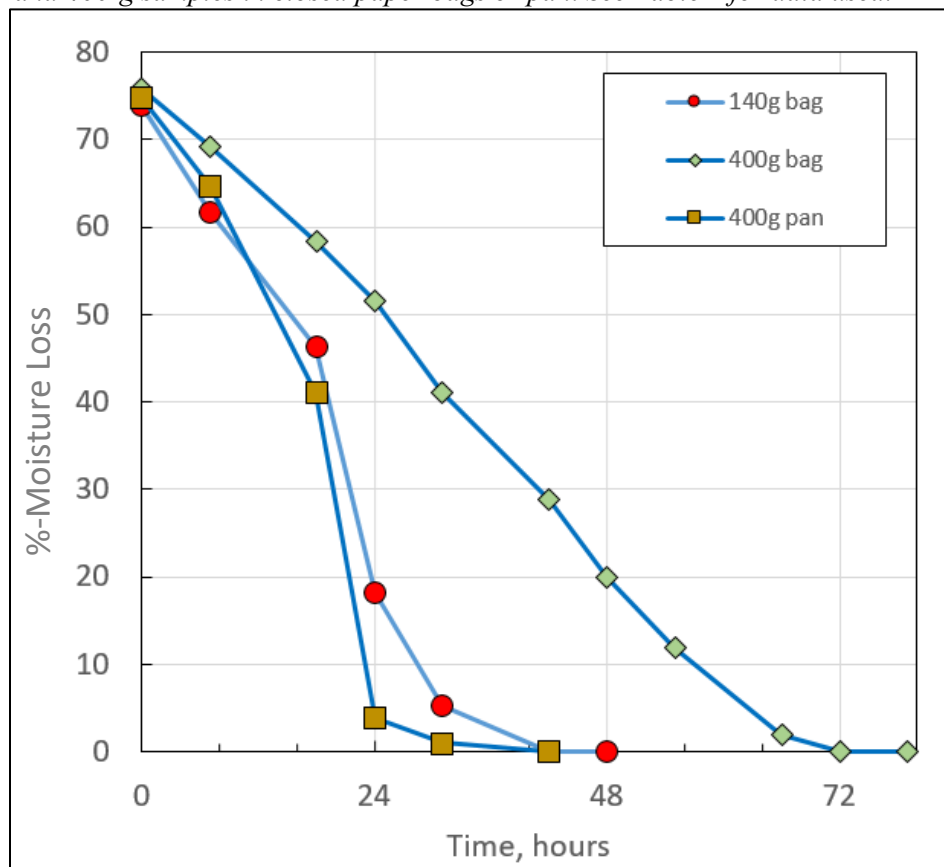
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For the examples shown above using a drying temperature of 60°C, 79 hours was needed to adequately dry a 400-g hemp sample using a closed paper bag, i.e., to reach constant weight defined as <5% weight change on two successive weighings indicating that no more moisture was being removed from the sample. Conversely, only 24 hours drying time was required for a 400-g sample dried using an open pan. Finally, 48 hours drying time was required for a 140-g sample dried in a closed paper bag.

The rate of moisture loss can be visualized by calculating %-Moisture Loss vs the last sample weighing and plotted vs weighing interval times as shown in Figure 1. Moisture losses of 0% are calculated for samples in which no more moisture is removed in drying and suggest drying is complete. Samples can be expected to have variable moisture content when removed from the oven and the material may in fact absorb some small amount of moisture prior to or during milling at laboratory ambient conditions.

Figure 1. Example Moisture-loss curves for field fresh hemp at 60°C for 140-g and 400-g samples in closed paper bags or pan. See Table 1 for data used.



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215 **5.0 Summary and Conclusions**

216 The goal of drying field-fresh hemp samples is not to remove all the moisture from the
217 samples but rather to reduce the moisture content of the plant material to a level appropriate for
218 grinding the material into a powdered analytical sample in preparation for analysis. The target
219 moisture content of the dried material is approximately 10-15% and is readily apparent when the
220 dried plant material disintegrates readily when crushed. Moisture content significantly higher
221 than this and the material will not mill properly. Moisture content less than 10% can lead to
222 change in cannabinoid profile and should also be avoided.

223 This guidance is intended to assist laboratories in developing and validating their internal
224 SOP for processing field fresh hemp to a dried state appropriate for milling and subsequent
225 preparation of an analytical sample for total THC analysis. To report percent total THC on a dry-
226 weight basis, an additional and separate analytical determination of moisture must be made on a
227 test portion of the sample taken at the same time a test portion is analyzed for total THC in
228 accordance with validated methods (see OMA 2018.11).

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231 **6.0 References**

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- 236 (3) Laboratory Testing Guidelines, US Domestic Hemp Production Program, Jan 15, 2021
- 237 (4) AOAC INTERNATIONAL, OMA Appendix K: Guidelines for Dietary Supplements and
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240 MS/MS method for detection and quantification of twelve cannabinoids”; C.R. McCurdy,
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